STEREOCONTROLLED SYNTHESIS OF 7-HYDROXY- AND 7-ACETOXY-PGI<sub>2</sub>: NEW STABLE PGI<sub>2</sub> ANALOGUES<sup>†</sup>

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Stable analogues of  $PGI_2$ , 7-hydroxy- and 7-acetoxy- $PGI_2$ , were synthesized from protected  $PGI_2$  methyl ester 1b via sulfoxides 6a, 6b through stereo-controlled sulfoxide-sulfenate rearrangement.

Since a part of limitations of  $PGI_2$  for clinical use is the hydrolysissensitive property due to its vinyl ether moiety,<sup>1</sup> stable  $PGI_2$  analogues have been attractive target molecules to be synthesized.<sup>2,3,4</sup> One of the efficient approaches to this end is to introduce an electron withdrawing group around the vinyl ether function. To date there exist a few reports concerning the synthesis of stable  $PGI_2$  analogues in this respect.<sup>3</sup> Previously we synthesized 5-PhS-PGI<sub>2</sub> 2a and 7-PhS- $\Delta^6$ -PGI<sub>1</sub> 3 and, in fact, introduction of the PhS-function in the vinyl ether double bond stabilized the vinyl ether linkage but lowered the biological activities as well.<sup>4</sup> To obtain chemically stable PGI<sub>2</sub> analogues retaining biological activity, we have explored another entry of stable PGI<sub>2</sub> analogues. We report here the stereocontrolled synthesis of 7-hydroxy- and 7-acetoxy-PGI<sub>2</sub> derivatives which were found to be more stable than PGI<sub>2</sub>.

In our previous study, <sup>4</sup> PGI<sub>2</sub> methyl ester 1a was subjected to react with PhSC1 to give vinyl sulfides 2a and 3, which would be formed partly *via* allyl sulfide intermediate 4a by way of the acid catalyzed rearrangement by HC1 produced during the reaction. When, in the presence of Et<sub>3</sub>N (3 eq.) to scavenge HC1, PhSC1 (1.5 eq.) was added to 1b in benzene at r. t., allyl sulfide 4b<sup>5</sup> could be obtained in 86% yield. <sup>6</sup> From the steric consideration of the *cis*bicyclo[3.3.0] system, it is reasonable that the phenylthio group approached from the less hindered  $\beta$ -side of the  $\Delta^5$ -double bond to form the episulfonium ion intermediate 5, whose C-7 proton was then eliminated to form allyl sulfide 4b. Thus, the configuration at C-5 of 4b was assigned to <u>R</u>. Interestingly, when a chloroform solution of 4b was treated with a catalytic amount of conc. H<sub>2</sub>SO<sub>4</sub> at r. t., the  $\Delta^6$ -double bond rapidly isomerized to give (SE)-sulfide 2b<sup>7</sup> in 83% yield without formation of the (5<u>2</u>)-isomer. This fact further supports the S<u>R</u>-configuration of 4b considering the acid catalyzed concerted allyl proton rearrangement initiated by the proton addition to C-7 from the less hindered



p-side. Oxidation of allyl sulfide 4b with m-chloroperbenzoic acid (1.2 eq.) in  $CH_2Cl_2$ -aqueous 0.05M NaHCO<sub>3</sub> gave an isomeric mixture of sulfoxides 6a and 6b in 53% yield, which could not be isolated as single isomers because of the rapid epimerization of the allyl sulfoxides *via* sulfenate ester  $\mathcal{I}$ , *i.e.* 6a  $\rightleftharpoons \mathcal{I} \rightleftharpoons 6b$ .<sup>8</sup> Treatment of the mixture of 6a and 6b with  $Et_2NH$  or  $(Me_2N)_3P$  (15 eq.) in THF (r. t., 12 h) gave rearranged alcohol  $g^9$  in 68% yield. The silyl protecting groups of g were removed with n-Bu<sub>4</sub>NF-Et<sub>3</sub>N (8 eq.) in THF (r. t., 2 h) to give (7S)-hydroxy-PGI<sub>2</sub> methyl ester 10 in 87% yield, which was hydrolyzed with NaOH-MeOH-H<sub>2</sub>O to give (7S)-hydroxy-PGI<sub>2</sub> sodium salt solution 12.

The stereochemistry of alcohol § was determined as follows. In the  ${}^{1}\text{H-nmr}$  spectrum the C-5 vinyl proton of § appeared at 4.38 ppm, while those of PGI<sub>2</sub> methyl ester 1a and its (5<u>E</u>)-isomer 1<u>3</u> have been reported to appear at 4.16 and 4.67 ppm, respectively.<sup>10</sup> Considering that the downfield shift by 0.22 ppm from PGI<sub>2</sub> methyl ester 1a was caused by the C-7 hydroxyl group,<sup>11</sup> the geometry of the

 $\Delta^5$ -olefin was assigned to <u>Z</u>. Furthermore, the <u>Z</u>-geometry of the  $\Delta^5$ -double bond was unambiguously determined on the basis of the assumption that (<u>5R</u>)-phenylsulfinyl group concertedly<sup>8</sup> migrated to the less hindered  $\beta$ -side of 7-position to form sulfenate ester <u>T</u> which lead to alcohol §.<sup>12</sup> Thus in order to form the (5Z)-isomer from sulfoxides <u>6a</u> and <u>6b</u>, the C-7 configuration of <u>8</u> must be <u>S</u>.

Treatment of 6a and 6b with  $Ac_20$ -pyridine (60°C, 2 h) gave acetate  $9^{\overline{13}}$  in 62% yield. Deprotection with n-Bu<sub>4</sub>NF-Et<sub>3</sub>N (10 eq.) in THF (r. t., 30 min) gave (7S)-acetoxy-PGI<sub>2</sub> methyl ester 11 which was hydrolyzed to give 12. The  $\Delta^5$ -olefin geometry was assigned to  $\underline{Z}$  as above from the <sup>1</sup>H-nmr study in which the C-5 vinyl proton of 9 appeared at 4.56 ppm, shifted downfield by 0.40 ppm from that of 1a. Since this rearrangement possibly proceeded *via* sulfenate ester  $\underline{7}$ , <sup>14</sup> the C-7 configuration was assigned to S as in the case of 8.

Interestingly, 7-oxygenated  $PGI_2$  derivatives 10 and 11 thus obtained were stable enough to be purified with normal silica gel preparative TLC without using base.<sup>15</sup> Moreover, the potency of the inhibition of platelet aggregation of 11 was maintained in a PH7.4 buffer solution for at least 4 h at r. t., while that of PGI<sub>2</sub> methyl ester 1a fell down to less than one tenth even at 0°C (4 h). The stability of these 7-oxygenated PGI<sub>2</sub> derivatives 10 and 11 are probably ascribed to the inductive effect<sup>16</sup> of the C-7 hydroxyl or acetoxy group. Analogue 11 had weak inhibitory activity on platelet aggregation and relatively higher hypotensive activity, but 10 and 12 did not show any of these activities.<sup>17</sup>

Acknowledgement; Authors are grateful to both Professor S. Ikegami, Teikyo Univ., and Professor R. Noyori, Nagoya Univ. for valuable suggestions in the structural determination of our reaction products.

## References and Notes

- + Prostaglandin Chemistry XV. For part XIV; see ref. 4.
- 1. Y. Chiang, A. J. Kresge, and M. J. Cho, Chem. Commun., 129 (1979) and references cited therein.
- For example, M. Shibasaki, Y. Torisawa, and S. Ikegami, Chem. Lett., 1247 (1980); W. Skuballa, Tetrahedron Lett., <u>21</u>, 3261 (1980); R. A. Johnson and E. G. Nidy, J. Org. Chem., 45, 3802 (1980); and references cited therein.
- H. Nishiyama and K. Ohno, Tetrahedron Lett., 3481 (1979); K. Ohno and H. Nishiyama, *ibid.*, 3303 (1979); J. Fried and J. Barton, Proc. Natl. Acad. Sci. USA, <u>74</u>, 2199 (1977); J. Fried, D. K. Mitra, M. Nagarajan, and M. M. Mehrotra, J. Med. Chem., <u>23</u>, 234 (1980); Schering AG, Ger. Offen. 2,801,846; Japan Kokai, 54-81257.
- T. Toru, K. Watanabe, T. Oba, T. Tanaka, N. Okamura, K. Bannai, and S. Kurozumi, Tetrahedron Lett., 21, 2539 (1980).

- 5.  ${}^{1}$ H-nmr (CCl<sub>4</sub>) & 3.65 (3H, s), 3.5-4.2 (3H, br), 4.6-4.9 (1H, br), 4.65 (1H, d, J=3Hz), 5.45 (2H, m); MS m/e 702 (M<sup>+</sup>).
- 6. All the yields indicated in this paper are isolated yields by preparative silica gel TLC or silica gel column chromatography.
- 7. Its  ${}^{1}$ H-nmr spectrum and TLC behavior completely coincided with those of (5E)-sulfide <u>2b</u> obtained previously.<sup>4</sup>
- P. Bickart, F. W. Carson, J. Jacobus, E. G. Miller, and K. Mislow, J. Am. Chem. Soc., <u>90</u>, 4869 (1968).
- 9. <sup>1</sup>H-nmr (CCl<sub>4</sub>) δ 3.66 (3H, s), 3.6-4.2 (2H, br), 4.14 (1H, s), 4.38 (1H, t, J=7Hz), 4.5-4.9 (1H, m), 5.5 (2H, m); MS m/e 610 (M<sup>+</sup>).
- R. A. Johnson, F. H. Lincoln, E. G. Nidy, W. P. Schnider, J. L. Thompson, and U. Axen, J. Am. Chem. Soc., <u>100</u>, 7690 (1978).
- 11. The following allyl hydroxyl group is known to cause the downfield shift of the *cis*- and *trans*-vinyl proton by 0.26 and 0.14 ppm, respectively; see Standard Proton Nmr Spectra Collection, No. 7926, Sadtler Research Laboratories, and E. J. Corey and H. Yamamoto, J. Am. Chem. Soc., <u>92</u>, 226 (1970).



- 12. D. A. Evans, and G. C. Andrews, Acc. Chem. Res., 7, 147 (1974).
- <sup>1</sup>H-nmr (CC1<sub>4</sub>) δ 2.01 (3H, s), 3.63 (3H, s), 3.6-3.9 (1H, m), 3.9-4.2 (1H, m), 4.56 (1H, t, J=7Hz), 4.5-4.8 (1H, m), 5.25 (1H, d, J=1Hz), 5.5 (2H, m); MS m/e 652 (M<sup>+</sup>).
- S. Yamagiwa, H. Sato, N. Hoshi, H. Kosugi, and U. Uda, J. Chem. Soc. Perkin I, 570 (1979).
- 15.  $PGI_2$  methyl ester <u>la</u> is usually purified by Florisil column chromatography using an eluent containing  $Et_3N$ .<sup>10</sup>
- 16. In the <sup>13</sup>C-nmr C-5 of acetate <u>11</u> (102.5 ppm) shifted downfield by 5.6 ppm from that of PGI<sub>2</sub> methyl ester <u>1a</u> (96.9 ppm)<sup>10</sup>, showing that the electron density of the  $\Delta^5$ -double bond of <u>11</u> is lower than that of <u>1a</u>.
- 17. Inhibitory activity on ADP-induced rabbit platelet aggregation of 11 was about one thousand times weaker (IC<sub>50</sub> = 8.2 µg/ml) than that of PGI<sub>2</sub> sodium salt, and 11 showed a transient fall of blood pressure at doses more than 1 µg/Kg which was about one hundred times weaker than that of PGI<sub>2</sub> sodium salt; submitted to J. Pharm-bio. Dyn.

(Received in Japan 4 January 1981)